New Fluorescent Monomers and Polymers Displaying an Intramolecular Proton-Transfer Mechanism in the Electronically Excited State (ESIPT). IV. Synthesis of Acryloylamide and Diallylamino Benzazole Dyes and Its Copolymerization with MMA

Leandra Franciscato Campo, Fabiano Severo Rodembusch, Valter Stefani

Universidade Federal do Rio Grande do Sul, Instituto de Química, Laboratório de Novos Materiais Orgânicos, Av. Bento Gonçalves, 9500. CP 15003 CEP 91501–970, Porto Alegre-RS, Brazil

Received 13 May 2005; accepted 28 June 2005 DOI 10.1002/app.22520 Published online 6 December 2005 in Wiley InterScience (www.interscience.wiley.com).

ABSTRACT: Six new fluorescent monomers were synthesized by reaction of 2-(5'-amino-2'-hydroxyphenyl)benzazole derivatives with acryloyl chloride and allyl bromide. UV–vis and steady-state fluorescence in solution were used to characterize its photophysical behavior. The monomers are fluorescent in the blue, green, yellow, and red region, with a large Stokes shift between 92 and 226 nm. A dual fluorescence ascribed to a conformational equilibrium in solution in the ground state dependent on the solvent po-

INTRODUCTION

ESIPT-exhibiting dyes are very attractive molecules, since they are highly fluorescent with a large Stokes shift, due to an excited-state intramolecular proton-transfer mechanism in the electronically excited state (Fig. 1).^{1–5} This phenomenon has implications in the UV-light stabilizers field,^{6,7} laser dyes,⁸ new polymeric materials,^{9–12} and also as fluorescent probes to labeling proteins.^{13,14}

The ESIPT mechanism is quite dependent on the solvent polarity,^{15–17} and many studies regarding this dependence,^{17–21} as well as, theoretical calculations involving the geometry of the conformers in solution^{22–27} have been made. In protic and/or polar solvents the enol-cis open conformer (E_{II}) can be stabilized by intermolecular hydrogen bond with the solvent.²⁸ This conformer is originated from the intramolecular hydrogen bond rupture between the hydrogen of the hydroxy group and the nitrogen in the third position followed by 180° rotation of the 2-hydroxyphenyl group under the C_2 – C_1 bond. In nonpolar solvents, additional enol-trans (E_{III}) con-

larity could be observed in the fluorescence emission spectra of the monomers. The radical polymerization of the monomers with methyl(methacrylate) allowed the production of fluorescent polymers in the blue–green region, with good optical and thermal properties. © 2005 Wiley Periodicals, Inc. J Appl Polym Sci 99: 2109–2116, 2006

Key words: copolymerization; dyes/pigments; fluorescence; photophysics; functionalization of polymers

formers in benzoxazoles and benzothiazoles (X = O and S, respectively) and enol-trans open (E_{IV}) in benzimidazoles (X = NH) could also exist (Scheme 1). All these conformers ($E_{II}-E_{IV}$) present normal relaxation and can compete with the keto tautomer responsible to the ESIPT mechanism.²⁹

Recently, we described new fluorescent MMA–benzazole dye copolymers with good optical and thermal properties.^{9,10} The synthesis and characterization of new acryloylamide and diallylamino derivatives and its radical polymerization with methyl(methacrylate), to produce new fluorescent polymers for optical application, were presented in this work.

EXPERIMENTAL

Materials

Acryloyl chloride and allyl bromide were purchased from Merck and used as received. The 2-(5'-amino-2'hydroxyphenyl)benzazoles **1a–c** were synthesized and purified according to a methodology previously described.⁹ Methyl(methacrylate) (MMA) (Aldrich) was purified before the polymerization reaction by passing on an activated alumina column. 2,2'-Azoisobutyronitrile (AIBN) was purchased from Merck and purified before use by recrystallization from methanol and maintained under vacuum. Silicagel 60 (Merck) was used for chromatographic column

Correspondence to: V. Stefani (vstefani@iq.ufrgs.br). Contract grant sponsors: CNPq, FAPERGS, CAPES.

Journal of Applied Polymer Science, Vol. 99, 2109–2116 (2006) © 2005 Wiley Periodicals, Inc.



Figure 1 ESIPT mechanism.

purifications. All the solvents were used as received or purified using standard procedures.

Instruments and methods

Infrared spectra were performed with a Mattson Galaxy Series FTIR 3000 in KBr. Melting points were measured with a Thermolyne apparatus and are uncorrected. ¹H and ¹³C NMR spectra were performed on a VARIAN model VXR-200 or INOVA-300, using tetramethylsilane (TMS) as the internal standard and DMSO- d_6 (Aldrich) or CDCl₃ (Merck) as the solvent, at room temperature. UV-vis absorption data were taken on a Shimadzu UV-1601PC spectrophotometer. Fluorescence spectra were measured with a Hitachi spectrofluorometer model F-4500. Spectrum correction was performed to enable measuring a true spectrum by eliminating instrumental response such as wavelength characteristics of the monochromator or detector, using Rhodamine B as a standard (quantum counter). Elemental analyses were performed using Perkin-Elmer model 240.

DSC analysis was performed with a Perkin–Elmer DSC-4. The standard operating range of the instrument was 50–300°C. Dry samples, 5–7 mg, were prepared in aluminum pan and sealed. A three-cycle method, from 50 to 300°C at a rate of 20°C min⁻¹, was used, with a nitrogen purge. TGA analysis was conducted with a Perkin–Elmer TGS-2 thermal gravimetric analyzer. Dry samples, 3–4 mg, were directly weighted into aluminum pans. A heating rate of 20°C min⁻¹ was maintained from 50 to 600°C. Size exclusion chromatography (SEC) was performed with a LDC Analytical Model Constametric 3200. Standard polystyrene was used as reference.

Synthesis of the monomer dyes

Synthesis of the 2-[5'-(N-Acryloyl)-2'hydroxyphenyl]benzazoles

The acryloyl derivatives $2\mathbf{a}-\mathbf{c}$ were prepared according to Scheme 2. To a solution of the corresponding benzazole $1\mathbf{a}-\mathbf{c}$ in chloroform, cooled at 5°C, was added dropwise a solution of an equimolar amount of acryloyl chloride in chloroform. After the addition, the mixture was stirred for 2 h to prepare $2\mathbf{a}$ and $2\mathbf{b}$ and 8 h to prepare $2\mathbf{c}$, cooled, and the resulting precipitate was filtered, washed with chloroform, dried at room temperature, and purified by column chromatography, eluted with chloroform.

2-[5'-(N-Acryloyl)-2'-hydroxyphenyl]benzoxazole (2a). Yield: 83%. m.p.: 252-253°C. Anal. Calcd for C₁₆H₁₂N₂O₃: C 68.56%, H 4.32%, N 9.99%. Found: C 68.27%, H 4.39%, N 9.76%. IR ν (cm⁻¹): 3296 (NH), 1661 (C=O), 1610 (C=C). ¹H NMR (200 MHz, DMSO d_6): δ (ppm) = 11.0 (s, 1H, OH); 10.2 (s, 1H, NH); 8.6 (d, 1H, $H_{6'}$, $J_{meta} = 2.7$ Hz); 8.5–7.8 (m, 2H, H_5 and H_6 or H₄ and H₇); 7.6 (dd, 1H, H_{4'}, $J_{\text{meta}} = 2.7$ Hz, $J_{\text{ortho}} = 9.0$ Hz); 7.5–7.4 (m, 2H, H_5 and H_6 or H_4 and H_7); 7.1 (d, 1H, $H_{3'}$, $J_{ortho} = 9.0$ Hz); 6.4 (dd, 1H, $CH_2 = CH - J_{cis}$ = 9.7 Hz, J_{trans} = 17 Hz); 6.3 (dd, 1H, CH₂=CH--, J_{gem} = 2.2 Hz, J_{trans} = 17 Hz); 5.8 (dd, 1H, CH₂=CH-, J_{gem} = 2.2 Hz, J_{cis} = 9.7 Hz). ¹³C NMR (50 MHz, DMSO- d_6): δ (ppm) = 163.8 (C=O); 152.6 (C₂); 151.4 (C_{2'}); 150 (C₈); 140.1 (C₉); 131.3 (C_{5'}); 130.8 (CH₂=CH-); 129.1 (CH₂=CH--); 125.4 (C₅ or C₆); 124.4 (C₅ or C₆); 123.9 $(C_{1'})$; 121.8 $(C_{4'})$; 120.5 (C_4) ; 120.3 $(C_{6'})$; 116.4 $(C_{3'})$; 110.8 (C₇).

2-[5'-(N-Acryloyl)-2'-hydroxyphenyl]benzothiazole (2b). Yield: 84%. m.p: 243–246°C. Anal. Calcd for $C_{16}H_{12}N_2O_2S$: C 64.85%, H 4.08%, N 9.45%. Found: C 64.66%, H 4.33%, N 9.41%. IR ν (cm⁻¹): 3271 (NH), 1658 (C=O), 1630 (C=C). ¹H NMR (200 MHz, CDCl₃): δ (ppm) = 11.4 (s, 1H, OH); 10.9 (s, 1H, NH); 8.5 (d, 1H, H_{6''}, J_{meta} = 3.0 Hz); 8.2–8.0 (m, 2H, H₅ and H₆ or H₄ and H₇); 7.7 (dd, 1H, H_{4'}, J_{meta} = 3.0 Hz, J_{ortho} = 9.0 Hz); 7.6–7.4 (m, 2H, H₅ and H₆ or H₄ and H₇); 7.0 (d,



Scheme 1



Scheme 2

1H, H_{3'}, $J_{\text{ortho}} = 9.0$ Hz); 6.4 (dd, 1H, CH₂=CH-, $J_{\text{cis}} = 9.0$ Hz, $J_{\text{trans}} = 17$ Hz); 6.3 (dd, 1H, CH₂=CH-, $J_{\text{gem}} = 2.4$ Hz, $J_{\text{trans}} = 17$ Hz); 5.8 (dd, 1H, CH₂=CH-, $J_{\text{gem}} = 2.20$ Hz, $J_{\text{cis}} = 9.0$ Hz). ¹³C NMR (50 MHz, CDCl₃): δ (ppm) = 171.4 (C₂); 163.0 (C=O); 153.5 (C₉); 151.4 (C_{2'}); 133.9 (C₈); 131.3 (C_{5'}); 130.0 (CH₂=CH-); 129.2 (CH₂=CH-); 125.8 (C₅ or C₆); 125.1 (C₅ or C₆); 123.9 (C_{1'}); 122.7 (C₇); 122.1 (C₄); 121.8 (C_{4'}); 120.3 (C_{6'}); 116.4 (C_{3'}).

2-[5'-(N-Acryloyl)-2'-hydroxyphenyl]benzimidazole (2c). Yield: 55%. m.p: 289–290°C. ¹H NMR (200 MHz, DMSO-*d*₆): δ (ppm) = 13.2 (s, 1H, OH or NH azolic); 12.9 (s, 1H, OH or NH azolic); 10.1 (s, 1H, NH amide); 8.5 (d, 1H, H₆', J_{meta} = 2.0 Hz); 7.7–7.6 (m, 2H, H₅ and H₆ or H₄ and H₇); 7.4 (dd, 1H, H₄', J_{meta} = 2.0 Hz, J_{ortho} = 9.0 Hz); 7.3–7.2 (m, 2H, H₅ and H₆ or H₄ and H₇); 7.0 (d, 1H, H₃', J_{ortho} = 9.0 Hz); 6.3 (dd, 1H, CH₂=CH–, J_{cis} = 10.0 Hz, J_{trans} = 27 Hz); 6.3 (dd, 1H, CH₂=CH–, J_{gem} = 5.0 Hz, J_{cis} = 10.0 Hz). ¹³C NMR (50 MHz, DMSO-*d*₆): δ (ppm) = 163.8 (C=O); 151.4 (C₂'); 141.5 (C₂); 137.9 (C₈ and C₉); 131.5 (C₅'); 130.2 (CH₂=CH–); 129.1 (CH₂=CH–); 123.9 (C₁'); 122.9 (C₅ and C₆); 121.8 (C₄'); 120.3 (C₆'); 116.4 (C₃'); 115.5 (C₄ and C₇).

Synthesis of the 2-[5'-(N,N-Dipropylamin-2-ene)-2'hydroxyphenyl]benzazoles

The diallylamino derivatives **3a–b** were prepared according to Scheme 2. To a solution of the corresponding benzazole **1a–c** in methanol was added an equimolar amount of sodium bicarbonate and stirred. To this solution was added allyl bromide (2:1 with respect to the benzazole dye) in methanol and it was allowed to react for 12 h at reflux. The dyes **3a–c** that precipitate into the reactional mixture were filtered, washed with chloroform, and dried at room temperature. The resulting products were purified by column chromatography, using chloroform as the eluent.

2-[5'-(N,N-Dipropylamin-2-ene)-2'-hydroxyphenyl]benzoxazole (3a).

Yield: 78%. m.p.: 93–95°C. Anal. Calcd for $C_{19}H_{18}N_2O_2$: C 74.49%, H 5.92%, N 9.14%. Found: C 74.10%, H 6.10%, N 8.95%. IR ν (cm⁻¹): 3471 (NH), 2973 (C—H), 1637–1631 (C=C). ¹H-NMR (200 MHz, CDCl₃): δ (ppm) = 10.90 (s, 1H, OH); 7.74–6.91 (m, 7H aromatic); 6.00–5.80 (m, 2H, —CH₂—CH=CH₂); 5.28–5.17 (m, 4H, —CH₂—CH=CH₂); 4.06–3.92 (m, 4H, —CH₂—CH=CH₂). ¹³C NMR (50 MHz, CDCl₃): δ (ppm) = 152.6 (C₂); 150.0 (C₈); 145.3 (C_{2'}); 140.9 (C₉); 137.6 (C_{5'}); 134.3 (—CH₂CH=CH₂); 125.4 (C₅ or C₆); 124.4 (C₅ or C₆); 124.6 (C_{1'}); 120.7 (C₄); 117.7 (C_{3'}); 114.9 (—CH₂CH=CH₂); 114.0 (C_{4'}); 113.0 (C_{6'}); 110.8 (C₇); 59.6 (—CH₂CH=CH₂).

2-[5'-(N,N-Dipropylamin-2-ene)-2'-hydroxyphenyl]benzothiazole (**3b**).

Yield: 82%. m.p.: 60–62°C. Anal. Calcd for C₁₉H₁₈N₂OS: C 70.78%, H 5.63%, N 8.69%. Found: C 70.58%, H 5.78%, N 8.83%. IR ν (cm⁻¹): 2926 (C—H), 1633–1591 (C=C). ¹H NMR (200 MHz, CDCl₃): δ (ppm) = 12.50 (s, 1H, OH); 8.10–7.10 (m, 7H aromatic); 6.20–6.00 (m, 2H, —CH₂—CH=CH₂); 5.42–5.35 (m, 4H, —CH₂—CH=CH₂); 4.07–4.04 (m, 4H, —CH₂ —CH=CH₂). ¹³C-NMR (50 MHz, CDCl₃): δ (ppm) = 171.4 (C₂); 153.5 (C₉); 145.3 (C_{2'}); 137.6 (C_{5'}); 134.0 (—CH₂CH=CH₂); 133.9 (C₈); 125.8 (C₅ or C₆); 125.1 (C₅ or C₆); 124.3 (C_{1'}); 122.7 (C₄ or C₇); 122.1 (C₄ or C₇); 117.5 (C_{3'}); 114.5 (—CH₂CH=CH₂); 114.0 (C_{4'}); 113.2 (C_{6'}); 59.3 (—CH₂CH=CH₂).

TABLE I AIBN, Fluorescent Dye, and MMA Amounts Used in the Polymerization

Copolymer	Dye	n _{dye}	$n_{\rm MMA} \ (\times 10^{-2})$	m _{AIBN} (mg)
Cop 2a	2a	$1.5 imes 10^{-5}$	9.3	12
Cop 2b	2b	$1.4 imes 10^{-5}$	9.3	12
Cop 2c	2c	$1.6 imes10^{-5}$	9.3	12
Cop 3a	3a	$9.8 imes 10^{-6}$	6.5	16
Cop 3b	3b	$6.2 imes 10^{-6}$	6.5	16
Cop 3c	3c	$8.1 imes 10^{-4}$	6.5	16

2-[5'-(N,N-Dipropylamin-2-ene)-2'-hydroxyphenyl]benzimidazole (3c).

Yield: 80%. m.p.: 234°C (decomp.). Anal. Calcd for $C_{19}H_{19}N_3O$: C 74.73%, H 6.27%, N 13.76%. Found: C 74.51%, H 6.00%, N 13.70%. IR ν (cm⁻¹): 2925 (C—H), 1626–1593 (C=C). ¹H NMR (200 MHz, DMSO-*d*₆): δ (ppm) = 7.90–7.10 (m, 7H aromatic); 5.92–5.77 (m, 2H, —CH₂—CH=CH₂); 5.32–5.20 (m, 4H, —CH₂—CH=CH₂); 4.07–3.90 (m, 4H, —CH₂—CH=CH₂). ¹³C NMR (50 MHz, DMSO-*d*₆): δ (ppm) = 145.5 (C₂·); 141.5 (C₂); 137.9 (C₈ and C₉); 137.0 (C₅·); 134.5 (CH₂CH=CH₂); 124.6 (C₁·); 122.9 (C₅ and C₆); 117.1 (C₃·); 115.4 (C₄ and C₇); 114.9 (CH₂CH=CH₂); 114.0 (C₄·); 113.5 (C₆·); 60.0 (N—CH₂CH=CH₂).

Polymer synthesis

The copolymers **Cop 2a–c** and **Cop 3a–b** were prepared by heating a solution of the fluorescent monomers in MMA. The amounts used for each polymer are presented in Table I. The initial temperature was 40°C for 2 days and then was increased up to 60°C. After 6 days, the samples were heated for 2 h at 70°C and subsequently the temperature was maintained at 80°C for 8 h.^{9,10} During the polymerization the temperature was maintained with an accuracy of ± 0.5 °C. The copolymers were purified by solubilization in chloroform and precipitation into cyclohexane (1:20 mL solvent/nonsolvent). The resulting fluorescent copolymers are presented in the Scheme 3.

RESULTS AND DISCUSSION

UV-vis and fluorescence characterization of monomer dyes

The UV–vis absorption and fluorescence emission spectra were made in dichloromethane (DCM) and ethanol. All experiments were performed at room temperature, in a concentration of $10^{-6}M$. The UV–vis absorption and fluorescence emission spectra are normalized. The relevant photophysical data are presented in Table II. As seen in Figure 2, the dyes **2a–c** present an absorption maximum (λ_{max}^{abs}) between 335 and 357 nm, with molar extinction coefficients ε values ($10^4 \text{ L mol}^{-1} \text{ cm}^{-1}$) in agreement with π – π * transitions ($S_0 \rightarrow S_1$). The absorption maximum of the dye **2c** presents a smooth dependence (~5 nm) on the solvent polarity. An intense band around 300 nm can be also observed, attributed to a charge-transfer mechanism due to the azole chromophore.²¹

The fluorescence emission spectra of the monomers 2a and 2c show the corresponding ESIPT band at 517 and 486 nm in ethanol, respectively, and at 517 and 495 nm in DCM. The monomer **2b** presented one main emission band at 547 nm in DCM, ascribed as the ESIPT band. However, in ethanol, an intense blueshifted band at 414 nm and the ESIPT band at 548 nm can be observed, indicating the conformational equilibrium in solution in the ground state. Usually, a dual fluorescence emission presents a band at higher wavelengths attributed to an excited keto tautomer (K^{*}), which arises from the enol-cis conformer (E_{I}) in the excited state, and a blue-shifted one due to the conformational forms, which present a normal relaxation,^{23,28} and the structure depends on the solvent polarity. In this way, for the dye **2b** in aprotic and/or



	0 v - v is and indorescence emission data of the dyes za-c and sa-c.								
Dye	Solvent	λ_{\max}^{abs} (nm)	$arepsilon_{\max} imes 10^{-4}$ $(1 \cdot \mathrm{mol}^{-1} \cdot \mathrm{cm}^{-1})$	$egin{aligned} & (\lambda^{\mathrm{em}}_{\mathrm{max}})_{\mathrm{enol}} \ & (\mathrm{nm}) \end{aligned}$	$\Delta\lambda_{ m ST}^{ m enol}$ (nm)	$egin{aligned} & (\lambda^{em}_{max})_{keto} \ & (nm) \end{aligned}$	$\Delta\lambda_{ m ST}^{ m keto}$ (nm)		
2a	Dichloromethane	345	1.32	_	_	517	172		
	Ethanol	345	0.58	_	_	517	172		
2b	Dichloromethane	357	0.82	_	_	547	190		
	Ethanol	357	0.89	414	42	548	191		
2c	Dichloromethane	340	1.20	—		495	155		
	Ethanol	335	0.75	_	_	486	151		
3a	Dichloromethane	394	0.54	480	86	591	197		
	Ethanol	397	0.66	474	77	584	187		
3b	Dichloromethane	405	0.51	470	65	631	226		
	Ethanol	408	0.58	500	92	_	92		
3c	Dichloromethane	387	0.25	433	46	565	178		
	Ethanol	373	0.55	436	63	545	172		

TABLE IIUV-Vis and fluorescence emission data of the dyes 2a-c and 3a-c.

low polar solvents, the conformational equilibrium is probably between the E_I and E_{III} conformers, which will emit fluorescence by normal relaxation. In protic solvents it is due to the conformers E_I and E_{II} .

In Figure 3 are presented the absorption and fluorescence emission spectra of the dyes 3a-c. All these dyes present a smooth dependence in the absorption maximum (λ_{max}^{abs}) on the solvent polarity. The dyes 3a-c presented the λ_{max}^{abs} at 397, 408, and 373 nm in ethanol, respectively, and 394, 405, and 387 nm in DCM, with molar extinction coefficients ε values (10⁴ L mol⁻¹ cm⁻¹) also in agreement with π – π * transitions. The intense band around 300 nm can also be observed, attributed to a charge transfer mechanism due to the azole chromophore.²¹

The fluorescence emission spectra of the monomers 3a-c in DCM show the corresponding ESIPT band at



Figure 2 Normalized absorption and fluorescence emission spectra of **2a**–**c**.



Figure 3 Normalized absorption and fluorescence emission spectra of **3a**–**c**.



Figure 4 Acryloyl copolymers under visible light (top) and UV radiation (bottom). [Color figure can be viewed in the online issue, which is available at www.interscience.wiley. com.]

591, 631, and 565 nm, respectively. Small blue-shifted bands can also be observed around 433–480 nm for **3a** and **3c**, indicating the conformational equilibrium in solution in the ground state, as already observed in the monomers **2a–c**. In the polar–protic solvent ethanol, the dual fluorescence can also be observed for **3a** and **3c**; however, with the blue-shifted band more intense, at 436–500 nm, in spite of the ESIPT band being above 545 nm. This behavior indicates that in ethanol, the conformers that present normal relaxation are in higher concentration in the ground state. The dye **3b** present one main emission band at 500 nm, ascribed to the ESIPT band.

Polymer characterization

The polymerization of the benzazolylvinylene dyes with MMA lead new fluorescent ESIPT polymers with the dyes covalently bonded into the polymer chain. The obtained fluorescent acryloyl copolymers **Cop 2a–c**, under visible light and UV radiation, are presented in Figure 4. The copolymers were analyzed by thermogravimetric analysis (TGA), differential scanning calorimetry (DSC), and size exclusion chromatography (SEC). The relevant data from the copolymers are summarized in the Table III. The fluorescent copolymers were also investigated by ¹H NMR and IR spectroscopy, where only the signals from the MMA polymer chain could be detected, as expected, since a very low concentration of the dyes where used in the polymerizations.⁹

Cop 2a–c and **Cop 3a–c**: IR *ν* (KBr pellets, cm⁻¹): 3020 (st C—H), 1740 (st C=O), 1240 (def C=O=C). ¹H NMR (200 MHz, CDCl₃) δ (ppm): 3.61 (3H, OCH₃), 1.87–1.81 (2H, CH₂), 0.85 (3H, CH₃). The **Cop 2a–c** present similar values of the glass transition temperature (T_g) when compared with PMMA. The **Cop 3a–c** presented higher T_g values, since the monomers are quite different than those used in the **Cop 2a–c**. In this way the polymer chain of **Cop 3a–c** seems to be more entangled than **Cop 2a–c**. The results from TGA show that the **Cop 3a–c** are more thermally stable than the **Cop 2a–c**, since values around 300°C were observed to the initial decomposition temperature (T_{di}) in relation to the **Cop 2a–c** $(T_{di} ~ 381^{\circ}\text{C})$.

The normalized absorption spectra and fluorescence emission of the **Cop 2a–c** in DCM $(10^{-5}M)$ are presented in Figure 5, and those of the monomers are also depicted for comparison. To these copolymers, the absorption maximum presented a small shift (1–4 nm) if compared to the monomers **2a–c**. The fluorescence emission spectra of the **Cop 2a–c** show no changes in its ESIPT band localization. However, the **Cop 2b–c** present additional blue-shifted bands at 421 and 400 nm, respectively, more intense to the Cop 2b in respect to the **Cop 2c**. The dual fluorescence emission, as already observed to the correspondent monomer, is probably due to the interaction of the phenolic OH with the polymer matrix, which contributes to the stabilization of the conformers that present normal relaxation.

The **Cop 3a–c** present a quite different photophysical behavior when compared with the monomers 3a-c (Fig. 6). The absorption maximum of the monomers could not be observed in the absorption spectra of the **Cop 3a–c**. All copolymers presented blue-shifted bands in the fluorescence emission spectra, indicating that the polymer matrix play a fundamental role in the photophysical behavior of the fluorescent dyes. The interaction of the phenolic OH with the MMA monomers seems to be more intense in the allyl than the acryloyl copolymer.

The fluorescent dye incorporation was determined using the copolymers' UV–vis data,^{9,12} through the benzazole molar extinction coefficient ε values at a

TABLE IIIRelevant Data of the Fluorescent Copolymers 2a-c and3a-c Obtained by SEC (M_n and M_w/M_n), DSC (T_g), TGA (T_{di} and T_{dr})

Sample	Т _{<i>g</i>} (°С)	Т _{di} (°С) 7	rdr(°C)	\bar{M}_n (×10 g mol ⁻¹)	\bar{M}_w/M_n
PMMA	125	240	381	469	1.4
Cop2a	116	260	385	560	2.2
Cop2b	127	290	383	698	2.1
Cop2c	126	305	387	594	2.1
Cop3a	132	295	461	915	1.6
Cop3b	120	317	458	188	3.7
Cop3c	130	302	461	282	2.6

PMMA was also prepared for better comparison.



Figure 5 Normalized absorption and fluorescence emission spectra of Cop 2a-c and monomers 2a-c.

specific wavelength (Lambert–Beer law). It was assumed that ε did not change when the benzazole was incorporated into the polymer chain. Values from 25 to 30% (w/w) [or 1.00–3.60% (mol/mol)] were found for the **Cop 2a–c**. Since the absorption maximum of the monomers could not be clearly observed in the absorption spectra of the **Cop 3a–c**, the dye incorporation in these samples could not be calculated.

CONCLUSIONS

New fluorescent acryloyl and allyl monomers were synthesized, purified until optical purity grade, and characterized by elemental analysis, ¹H NMR, ¹³C NMR, IR, UV–vis, and steady-state fluorescence spectroscopy. The monomers are fluorescent in the blue-green–yellow–red region and present a Stokes shift between 92 and 226 nm. The absorption maximum dependence on the solvent polarity and the dual flu-



Figure 6 Normalized absorption and fluorescence emission spectra of Cop 3a-c and monomers 3a-c.

orescence indicated a conformational equilibrium in solution in the ground state. The emission at higher wavelength (ESIPT band) is due to an excited keto tautomer which arises from an enol-cis (E_1) conformer in the excited state. The blue-shifted bands are due to conformational forms with a normal relaxation. The new monomers are found to be very sensitive to solvent polarity. All the synthesized monomers were totally soluble in MMA and were used to produce new fluorescent polymers. The resultant copolymers are transparent in the visible light and present fluorescence when illuminated with UV radiation. The new monomers and polymers showed to be very attractive to be used as new fluorescent materials.

References

- 1. Arnaut, L. G.; Formosinho, S. J. J Photochem Photobiol A 1993, 75, 1.
- 2. Formosinho, S. J.; Arnaut, L. G. J Photochem Photobiol A 1993, 75, 21.
- 3. Elguero, J.; Katritzky, A. R.; Denisko, O. V. Adv Heterocycl Chem 2000, 76, 1.
- Minkin, V. I.; Garnovskii, A. D.; Elguero, J.; Katritzky, A. R.; Denisko, O. V. Adv Heterocycl Chem 2000, 76, 157.
- 5. Doroshenko, A. O.; Posokhov, E. A.; Verezubova, A. A.; Ptyagina, L. M. J Phys Org Chem 2000, 13, 253.
- Catalán, J.; Fabero, F.; Guijarro, M. S.; Claramunt, R. M.; Santa Maria, M. D.; Foces-Foces, M. C.; Cano, F. H.; Elguero, J.; Sastre, R. J Am Chem Soc 1990, 112, 747.
- Kuila, D.; Kwakovszky, G.; Murphy, M. A.; Vicare, R.; Rood, M. H.; Fritch, K. A.; Fritch, J. R. Chem Mater 1999, 11, 109.
- 8. Uzhinov, B. M.; Druzhinin, S. I. Russ Chem Rev 1998, 67, 123.

- Campo, L. F.; Corrêa, D. S.; Araújo, M. A.; Stefani, V. Macromol Rapid Commun 2000, 21, 832.
- Rodembusch, F. S.; Leusin, F. P.; Bordignon, L. B.; Gallas, M. R.; Stefani, V. J. Photochem Photobiol A 2005, 173, 81.
- Rodembusch, F. S.; da Silveira, N. P.; Samios, D.; Campo, L. F.; Stefani, V. Mol Cryst Liq Cryst 2002, 374, 367.
- 12. Rodembusch, F. S.; da Silveira, N. P.; Samios, D.; Campo, L. F.; Stefani, V. J Polym Sci Part B: Polym Phys 2003, 41, 341.
- Rodembusch, F. S.; Leusin, F. P.; Medina, L. F. C.; Brandelli, A.; Stefani, V. Photochem Photobiol Sci 2005, 4, 254.
- Holler, M. G.; Campo, L. F.; Brandelli, A.; Stefani, V. J Photochem Photobiol A 2002, 149, 217.
- 15. Nagaoka, S.; Itoh, A.; Mukai, K. J Phys Chem 1993, 97, 11385.
- 16. Kasha, M.; McMorrow, D. J Am Chem Soc 1983, 105, 5133.
- 17. Das, K.; Sarkar, N.; Gosh, A. K.; Majumdar, D.; Nath, D. N.; Bhattacharyya, K. J Phys Chem 1994, 98, 9126.
- Santra, S.; Krishnamoorthy, G.; Dogra, S. K. Chem Phys Lett 1999, 311, 55.
- 19. Dogra, S. K.; Balamurali, M. M. J Photochem Photobiol A 2002, 154, 81.
- 20. Klymchenko, A. S.; Pivovarenko, V. G.; Demchenko, A. P. J Phys Chem A 2003, 107, 4211.
- 21. LeGourriérec, D.; Kharlanov, V. A.; Brown, R. G.; Rettig, W. J Photochem Photobiol A 2000, 130, 101.
- 22. Purkayastha, P.; Chattopadhyay, N. Int J Mol Sci 2003, 4, 335.
- 23. Santra, S.; Dogra, S. K. Chem Phys 1998, 226, 285.
- 24. Forés, M.; Duran, M.; Solà, M.; Orozco, M.; Luque, F. J. J Phys Chem A 1999, 103, 4525.
- 25. Premvardhan, L.; Peteanu, L. Chem Phys Lett 1998, 296, 521.
- Forés, M.; Duran, M.; Solà, M.; Adamowicz, L. J Phys Chem A 1999, 103, 4413.
- 27. Purkayastha, P.; Chattopadhyay, N. Phys Chem Chem Phys 2000, 2, 203.
- 28. Sinha, H. K.; Dogra, S. K. J Chem Soc Perkin Trans 2 1987, 1465.
- 29. Roberts, E. L.; Dey, J.; Warner, I. M. J Phys Chem A 1997, 101, 5296.